## Synthesis of Carthamin Acetate, the Red Pigment in Safflower Petals

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Carthamin acetate, (3S,3'S)-1-[5-(*p*-acetoxycinnamoyl)-3-*C*-( $\beta$ -D-2",3",4",6"-tetra-*O*-acetylglucopyranosyl)-3,4-dihydroxy-2,6-diketo]cyclohex-4-enylidene-1'-[5'-(*p*-acetoxycinnamoyl)-3'-*C*-( $\beta$ -D-2"',3"',4"',6"'-tetra-*O*-acetylglucopyranosyl)-2', 3',4'-trihydroxy-6'-keto]cyclohexa-1',4'-dienylmethane **7a** and its (3*R*,3'*R*)-epimer **7b** were synthesized via the *C*-glycosylation of 2-acetyl-1,3,4,5-benzenetetrol using acetobromoglucose in the presence of sodium hyride, followed by aldol condensation with *p*-hydroxybenzaldehyde after methyl-protection of the enolic hydroxy group, and subsequent dimerization with triethyl orthoformate after demethylation, in a total yield of 0.6%. Their absolute configurations were determined by X-ray analysis of the key intermediate **4b**.

The structure of carthamin, the red pigment in the flowers of the Safflower plant (*Carthamus tinctorius* L.), was revised as **1** by H. Obara and J. Onodera in 1979,<sup>1</sup> based on some previous structural proposals of some analogues,<sup>2</sup> and has now been proved by the synthesis of model compounds (Figure 1).<sup>3</sup> The absolute configuration of the two-carbinol carbons at the 3- and 3'-positions to which D-glucose is linked has recently been compared with some chiral model compounds and determined to be *S*, *S*.<sup>4</sup> This red pigment, which is used as a food colorant and as rouge in the cosmetic industry in Japan, is a highly oxidized and complex keto–enol mixture of *C*-glucosylquinochalcone dimers and is strongly acidic. Therefore, it has not been reported to date that the acetate **7a** is derived from **1**.<sup>5</sup> Herein, we wish to report the first total synthesis of carthamin acetate **7a** and its epimer **7b**.





The synthesis of the key intermediate, cyclohexadienone, attached to a sugar moiety is very difficult and still remains, although the synthesis of the model compound in which the sugar moiety is replaced by a methyl group was achieved.<sup>3d,4</sup> By *C*-glycosylation of 2,6-diacetyl-1,3,4,5-benzenetetrol with acetobromoglucose in the presence of sodium hyride, the synthesis of *C*- $\beta$ -glucosylcyclohexadienone has been reported in a yield of 12%.<sup>6</sup> However, since the subsequent mono-deacetylation step failed, we attempted the direct *C*-glycosylation of 2-acetylbenzene-1,3,4,5-tetrol. When 3.5 equivalents of sodium hydride were used, the key intermediate, *C*- $\beta$ -D-glucopyra-



Reagents and conditions: a) NaH, per-O-acetylglucopyranosyl bromide, in DMSO. b) 1. CH<sub>2</sub>N<sub>2</sub> in EtOAc, 2. Ac<sub>2</sub>O/pyridine, 11% (from 2, 4a:4b=1.1: 1.0). c) 1. NaOMe in MeOH, 2. Dowex 50 W(H<sup>+</sup>). d) 1. *p*-hydroxybenz-aldehyde, piperidine, at 80 °C, 2. Ac<sub>2</sub>O/pyridine, (5a: 42%, 5b: 45%). e) trimethylsilyl chloride/NaI (10 equiv) in CH<sub>3</sub>CN at rt. f) NaH, in CH(OEt)<sub>3</sub>, (7a: 26%, 7b: 25%, from 5a and 5b).

nosylcyclohexadienone was obtained. After protection of the enol-hydroxy function of 3 with diazomethane, in order to prevent an aldol reaction between the 1,3-diketone and isomerization,<sup>7</sup> followed by acetylation with Ac<sub>2</sub>O/pyridine, the desired C- $\beta$ -D-glucopyranosylcyclohexadienone acetate 4 was readily isolated from the diastereomixture (4a:4b=1.1:1.0),<sup>8</sup> in a low yield of 11% from 2. Fortunately, by means of an X-ray crystal structure analysis of 4b, the absolute configuration of 4b was determined to be R (Figure 2), and, thus, its epimer 4a to be S. Compounds 4a and 4b were deacetylated with NaOMe, followed by aldol condensation with p-hydroxybenzaldehyde in the presence of piperidine at 80 °C and then re-acetylated with Ac<sub>2</sub>O/pyridine to give acetylated C-β-glucosylquinochalcones 5a (3S) and 5b (3R) in 50 and 37% yield respectively.<sup>8</sup> Each chalcone 5a and 5b was next subjected to demethylation by treatment with trimethylsilyl iodide in CH<sub>3</sub>CN at room temperature for 4 h to afford 6a and 6b. Each activated monomer acetate 6a and 6b was then dimerized by treatment with triethyl orthoformate in the presence of a catalytic amount of sodium hydride to give the reddish colored carthamine acetate 7a (3S, 3'S) and its epimer 7b (3R, 3'R) in 26 and 25% yield from 5a and **5b**, respectively.<sup>8</sup> The electronic spectra of **7a** and **7b** showed an absorption maximum (red) at around 520 nm similar to that of carthamin 1.9 A comparison of the CD spectra of 7a



**Figure 2.** ORTEP drawing of **4b**.  $P2_1$  (monoclinic), a=10.70 (1), b=33.02 (4), c=7.962 (5) Å,  $\beta=90.51$  (8)°, V=2811 (4) Å<sup>3</sup>, Z=4,  $D_{\rm C}=1.310$  g/cm<sup>3</sup>, v=9.31 cm<sup>-1</sup>, Cu K $\alpha(\lambda=1.54178$  Å), Crystal dimensions  $0.20 \times 0.05 \times 0.30$  mm,  $R(R_{\rm W})=0.088$  (0109).



Figure 3. CD spectra of 7a, 7b, and 1.

and **7b** with that of **1** revealed that the curve for **7a** was analogous to that of **1**, on the contrary, that of **7b** yielded a curve which was roughly opposite in sign (Figure 3). Thus, the synthesis of carthamin acetate was achieved for the first time.

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## **References and Notes**

- a) H. Obara and J. Onodera, *Chem. Lett.*, **1979**, 201. b) Y. Takahashi, N. Miyasaka, S. Tasaka, I. Miura, S. Urano, M. Ikura, K. Hikichi, T. Matsumoto, and M. Wada, *Tetrahedron Lett.*, **23**, 5163 (1982).
- a) C. Kuroda, Nippon Kagaku Kaishi, 51, 237 (1930). b) C. Kuroda,
   J. Chem. Soc., 1930, 752. c) T. R. Seshadori and R. S. Thakur,
   Curr. Sci., 29(2), 54 (1960).
- 3 a) J. Onodera, T. Saito, and H. Obara, *Chem. Lett.*, **1979**, 1327. b)
  H. Obara, J. Onodera, and F. Shirasaki, *Chem. Lett.*, **1980**, 1095. c)
  H. Obara, J. Onodera, S. Abe, and T. Saito, *Bull. Chem. Soc. Jpn.*, **53**, 289 (1980). d)
  H. Obara, S. Namai, and Y. Machida, *Chem. Lett.*, **1986**, 495.
- 4 S. Sato, H. Obara, T. Kumazawa, J. Onodera, and K. Furuhata, *Chem. Lett.*, **1996**, 833. In this paper, the direct methylation of 2acetyl-1,3,4,5-benzenetetrol with methyl iodide, gave the desired cyclohexadinenone in 56% yield.

- 5 A typical acetylation of 1 with  $Ac_2O$ /pyridine or acid did not give the acetate 7a.
- 6 H. Obara, Y. Namai, and Y. Machida, Chem. Lett., 1984, 1397.
- 7 H. Obara, Y. Machida, S. Namai, and J. Onodera, *Chem. Lett.*, 1985, 1393.
  - **4a**: colorless syrup.  $[\alpha]_D^{25}$  –22.0° (*c* 1.00 CHCl<sub>3</sub>). EI-MS (*m*/*z*) 570 (M<sup>+</sup>), 510, 468, 331, 268, 167. <sup>1</sup>H NMR (500MHz, CDCl<sub>3</sub>)  $\delta$  = 1.99 (6H, s, OAc × 2), 2.02, 2.05 and 2.11 (each 3H, s, OAc × 3), 2.55 (3H, s, Ac), 3.56 (1H, ddd, J = 9.5, 2.0, and 6.5 Hz, H-5'), 3.78 (3H, s, OCH<sub>3</sub>), 3.88 (1H, dd, J = 6.5 and 13.5 Hz, H-6'a), 3.98 (1H, d, J = 9.5 Hz, H-1'), 4.07 (1H, dd, *J* = 2.0 and 13.5 Hz, H-6'b), 4.95 (1H, t, J = 9.5 Hz, H-4'), 5.11 (1H, t, J = 9.5 Hz, H-3'), 5.17 (1H, t, J = 9.5 Hz, H-2'), 5.58 (1H, s, olefinic H), 18.37 (1H, s, OH). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta = 20.1-21.0$  (OCC<u>CH<sub>3</sub></u> × 5), 27.0 (CC<u>OCH<sub>3</sub></u>), 56.9 (OCH<sub>3</sub>), 62.1, 67.8, 69.0, 69.9, 74.5, and 79.8 (sugar moiety), 92.0 (C4), 99.1 (C2), 108.3 (C5), 168.7-172.5 (OCCCH<sub>3</sub>) ×5), 188.7 (C3), 189.6 (C7), 198.6 (C1). 4b: colorless prism. mp 176–178 °C. EIMS (m/z) 570 (M<sup>+</sup>), 331, 268, 196, 167. [ $\alpha$ ]<sub>D</sub><sup>25</sup>  $-31.9^{\circ}$  (c 1.00 CHCl<sub>3</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta = 2.00$  (9H, s, OAc × 2.06 and 2.09 (each 3H, s,  $OAc \times 3$ ), 2.57 (3H, s, Ac), 3.56 (1H, ddd, J = 3.5, 7.5, and 9.5 Hz, H-5'), 3.81 (3H, s,  $OCH_3$ ), 4.02 (1H, dd, J = 7.5 and 13.5 Hz, H-6'a), 4.02 (1H, d, J = 9.5 Hz, H-1'), 4.04 (1H, dd, J = 3.5 and 13.5 Hz, H-6'b), 4.92 (1 H, dd, J = 9.5 Hz, H-(1), 5.16 (1H, t, J = 9.5 Hz, H-3'), 5.41 (1H, t, J = 9.5 Hz, H-2'), 5.55 (1 H, s, olefinic H-5), 18.31 (1H, s, OH). 5a: pale-yellow syrup. TLC:  $R_{\rm f} = 0.38$  (toluene–EtOAc–AcOH = 5:2:0.5).  $[\alpha]_{\rm D}^{24}$ –6.23° (c 120:  $R_1 = 0.50$  (challe Elove From = 0.250.5).  $[G_{10}]_{D} = 0.25$  (c 0.995 CHCl<sub>3</sub>). FABMS (*m/z*) 717(M+H). <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>)  $\delta = 2.01, 2.09, 2.11, 2.16, and 2.23 (each 3H, s, OAc × 5),$ 2.42 (3H, s, ArOAc), 3.68 (1 H, ddd, J = 2.0, 6.4, 9.8 Hz, H-5) 3.89 (3H, s, OCH<sub>3</sub>), 3.97 (1H, dd, J = 6.4 and 12.2 Hz, H-6'a), 4.10 (1H, d, J = 9.8 Hz, H-1'), 4.14 (1H, dd, J = 2.0 and 12.2 Hz, H-6'b), 5.05 (1H, t, J=9.8 Hz, H-4'), 5.18 (1H, t, J = 9.8 Hz, H-3'), 5.24 (1H, t, J = 9.8 Hz, H-2'), 5.75 (1H, s, olefinic H), 7.31 and 7.76(each 2H, J = 8.5 Hz, p-substituted ArH  $\times$  4), 7.79 and 8.31 (each 1H, J = 15.6 Hz, trans-vinyl H × 2), 18.64 (1H, s, OH). <sup>13</sup>C NMR  $(100 \text{ MHz}, \text{CDCl}_3) \delta = 56.9, 62.1, 67.8, 69.0, 74.6, 76.1, 77.4, 79.7, 80.7, 100.9, 107.5, 120.2 (× 2), 130.0, 132.7, 144.1, 152.5, 184.5,$ 189.2, 191.7, and OAc × 6. **5b**: pale-yellow crystal. mp = 110 °C. TLC:  $R_{\rm f} = 0.42$  (toluene–EtOAc–AcOH = 5:2:0.5).  $[\alpha]_{\rm D}^{24}$  –49.8° (c 1.02 CHCl<sub>3</sub>). FABMS (m/z) 717 (M+H). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  = 1.92, 1.99, 2.00, 2.06, and 2.11 (each 3H, s, OAc × 4), 2.32 (3H, s, ArOAc), 3.56 (1H, ddd, J = 2.5, 5.0, and 9.0 Hz, H-5'), 3.81 (3H, s, OCH<sub>3</sub>), 4.01 (1H, dd, J = 5.0 and 12.5 Hz, H-6'a), 4.03 (1H, d, J = 9.0 Hz, H-1'), 4.06 (1H, dd, J = 2.5 and 12.5 Hz, H-6'b), 4.95 (1 H, t, J = 9.0 Hz, H-4'), 5.16 (1H, t, J = 9.0 Hz, H-3'), 5.42 (1H, t, J = 9.0 Hz, H-2'), 5.61 (1H, s, olefinic H), 7.16 and 7.71 (each 2H, d, J = 8.5 Hz, *p*-substituted ArH × 4), 7.90 and 8.10 (each 1H, d, J = 16.0 Hz, trans-vinyl H × 2), 18.40 (1H, s, OH). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ = 57.1, 61.5, 67.5, 68.1, 74.6, 75.7, 77.5, 78.8, 80.6, 100.4, 107.8, 122.0, 122.1, 130.2, 132.6, 144.0, 152.5, 184.3, 187.3, 191.5, and OAc  $\times$  6. **7a**: red solid. TLC:  $R_{\rm f} = 0.47$  $(CHCl_3-MeOH = 8:1)$ . ESIMS (negative) (m/z) 1413.4 (M - H), 729, 701. UV/vis (EtOH) (log ε) 230 (4.53), 354 (4.47), 529 (4.72) m. CD (EtOH) λ<sub>ext</sub>(Δε) 275 (-13.9), 322 (+3.8), 334 (+4.4), 325 (+3.2) (+3.2), 322 (+5.8), 384 (-6.06), 540 (+5.71) nm. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ , at 50 °C) δ = 1.91 (12H, s, OAc × 4), 1.94 (12H, s, OAc × 4), 2.02 (6H, s, OAc × 2), 2.28 (6H, s, ArOAc × 2), 3.86 (2H, ddd, J = 9.0, 5.5, and 3.5 Hz, H-5" × 2), 3.95 (4H, m, H-6"a, b × 2), 4.41 (2H, d, J = 9.0 Hz, H-1" × 2), 4.80 (2H, t, J = 9.0 Hz, H-4" × 2), 5.25 (2H, t, J = 9.0 Hz, H-2" × 2), 7.22 (4H, d, J = 8.5 Hz, *p*-substituted ArH × 4), 7.70 (4H, d, J =8.5 Hz, *p*-substituted ArH × 4), 7.78 (2H, d, J = 15.9 Hz, *trans*-vinyl H × 2), 8.13 (2H, d, J = 15.9 Hz, *trans*-vinyl H × 2), 8.23 (1H, s, =CH-), 19.22 (2H, s, OH× 2). 7b: red solid. TLC:  $R_f = 0.25$  $(CHCl_3-MeOH = 8:1)$ . ESIMS (negative) (m/z) 1413 (M - H), 729, 701. UV/vis (EtOH) (log ε) 230 (4.44), 352 (4.43), 523 (4.71) nm. CD (EtOH)  $\lambda_{ext}(\Delta\epsilon)$  268 (+18.3), 316 (-13.6), 400 (-5.05), 542 (-2.44) nm. <sup>1</sup>H NMR (500 MHz, DMSO- $d_6$ , at 40 °C)  $\delta$  = 1.81, 1.90, 1.93, 1.99, and 2.05 (each 6H, s,  $OAc \times 10$ ), 2.29 (6H, s, ArOAc × 2), 3.77 (2H, d, J = 11.8 Hz, H-6"a × 2), 3.83 (2H, m, H- $5'' \times 2$ ), 3.91 (2H, dd, J = 11.8 and 4.9 Hz, H-6"b), 4.45 (2H, d, J =8.5 Hz, H-1" × 2), 4.77 (2H, t, J = 9.4 Hz, H-4" × 2), 5.22 (4 H, m, tuted ArH × 8), 7.75 and 7.84 (each 2H, d, J = 8.5 Hz, p-substituted ArH × 8), 7.75 and 7.84 (each 2H, d, J = 16.4 Hz, trans-vinylHere AIT  $\times$  6), 7.75 and 7.65 (etc. 213, e. C. 214) H × 4), 8.16 (1H, br. s. =CH-), 19.2 (2H, s. OH × 2). Natural carthamin 1: UV/vis (EtOH) (log  $\epsilon$ ) 244 (4.33), 373 (4.48),
- 9 Natural carthamin 1: UV/vis (EtOH) (log  $\epsilon$ ) 244 (4.33), 373 (4.48), 515 (4.99) nm. CD (EtOH)  $\lambda_{ext}(\Delta\epsilon)$  218 (–21.4), 271 (–24.6), 321 (+11.3), 361 (+4.02), 400 (–4.2), 509 (+7.61) nm.